



## BIOCHEMICAL EVALUATION OF ANTITUMOR ACTIVITY OF IRRADIATED CITRUS PECTIN.

Omayma A.R. Abou Zaid<sup>a</sup>, El-Batal, A.I.<sup>b</sup> and Effat, S.I.<sup>a</sup>

<sup>a</sup> Biochemistry Department, Fac. Vet. Med., Benha University, Egypt. <sup>b</sup> Drug Radiation Research Department, Biotechnology Division, National Center for Radiation Research and Technology (NCRRT), Atomic Energy Authority, Egypt.

### ABSTRACT

This study was carried out to evaluate the antitumor activity of citrus pectin (CP) and irradiated citrus pectin (Irr. CP) on 60 female mice and weighting 20-25 g. the mice were divided into four equal groups of 15 mice. Group 1: Non-tumor bearing mice (NTBM), Group 2: Tumor bearing mice (TBM), Group 3: TBM-treated with citrus pectin orally (3.3g/kg.b.w./day) for 4 weeks and Group 4: TBM-treated with Irr.CP orally (3.3 g /kg b.wt./day) for 4 weeks. Blood samples were collected from all animals after 2 and 4 weeks from the onset of treatment and processed directly for determination of SOD, GPx, and CAT activities in addition to MDA, GSH, and NO(x) concentrations. Also, AST, ALT and GGT activities as well as urea and creatinine concentrations were also determined in blood. The obtained results revealed a significant increase in MDA , No(x) level , CAT activity and in liver and kidney functions ratios, a marked significant depletion in GSH content, GPx and SOD activity in the blood of tumor bearing mice compared to control. Contrary results obtained in TBM treated with CP and Irr.CP. So, these compounds have potential benefits in cancer treatment.

**KEY WORDS:** antitumor activity, Mice, Pectin

(BVMJ 23(1): 11-18, 2012)

### 1. INTRODUCTION

Cancer is considered one of the major causes of mortality in the world. Despite the recent advances in science, cancer has not been cured yet. It is estimated that by 2020 there will be 16 million new cancer cases every year [1]. The target of much research has been on the discovery of natural and synthetic compounds that can be used in the prevention and/or treatment of cancer. Natural products of either plant or animal origin that exhibited antitumor activity have been discovered [2].

Pectin is a highly complex branched polysaccharide fiber rich in galactoside residues and present in all plant cell walls [3]. Ordinary pectin isolated from citrus fruits has high molecular weight and can be modified resulting in shorter, less complex molecules. These shorter

carbohydrate chains, dissolve more readily in water and are better absorbed and utilized by the body than ordinary, long-chain pectin [4]. Moreover, pectin may be modified by physical means. Such physical means include, but are not limited to heat, cold, freeze/thaw, irradiation, shear, ultra-high shear. Gamma irradiation is a useful physical treatment for depolymerizing pectin. Gamma irradiation leads degradation of polymer molecules by the free radical formed [5]. Irradiation induced degradation has been applied to preparing low molecular weight pectin [6] product with increased amount of monosaccharaides.

Citrus Pectin and modified Citrus pectin have been found to exhibit antimutagenic activity and inhibit cancer metastasis and proliferation, with no evidence of toxicity

or other serious side effects [7,8]. Accordingly, this study was performed to investigate the biochemical effect of citrus pectin and irradiated citrus pectin in experimentally induced tumor in female mice.

## 2. MATERIAL AND METHODS

### *Animals:*

Female Swiss albino mice weighting 20-25 g used in this study were obtained from Laboratory Animals Research Center, Faculty of Veterinary Medicine, Zagazig University, Egypt. The animals were housed in separated metal cage 10-15 per cage and kept at a under the same constant environmental and nutritional condition throughout the period of investigation, water was supplied *ad-Libitum*, in the special lab, animal room, in Fac. Vet. Med., Benha University.

### *Ehrlich Ascites Carcinoma Cells:*

A line of Ehrlich Ascites Carcinoma (EAC) cells was supplied from National Cancer Institute, Cancer Biology Department, Egypt.

### *Tumor induction:*

Solid tumors were induced by intramuscular inoculation of each mouse with 0.2 ml of EAC, which contained  $2.5 \times 10^6$  viable EAC cells, in the right thigh of the lower limb of each mouse. Mice with a palpable solid tumor, its diameter was 10mm<sup>3</sup>, that developed within 10 days after inoculation were used in the study.

### *Chemicals:*

Citrus Pectin purchased from El-Goumhouria Co. for trading chemicals, medicines and medical appliances, Egypt. All chemical and kits purchased from Segma (USA).

### *Nutraceuticals preparation:*

#### *Preparation of 4% citrus pectin:*

Four grams of citrus pectin were dissolved in 100ml distilled water.

#### *Preparation of irradiated citrus pectin:*

Prepare 4% citrus pectin solution in distilled water. This solution was irradiated with a dose of 5 kGy (kilo Gray) gamma radiation. The irradiation process was performed at National Center of Radiation Research and Technology (NCRRT, Cairo), Egypt.

#### *Experimental design:*

Sixty female mice were divided into 4 groups each one contains 15 mice placed in individual cages and classified as follows:

1. Group (1): Served as negative control and orally received saline (NTBM: Non-tumor bearing mice).
2. Group (2): Tumor bearing mice without any treatment served as positive control group (TBM) for 4 weeks.
3. Group (3): Tumor bearing mice received citrus pectin orally at a dose level of 3.3gm /Kg body weight/day (TBM<sub>(CP)</sub>) for 4 weeks
4. Group (4): Tumor bearing mice received Irr.CP orally at a dose level of 3.3gm /Kg body weight /day (TBM<sub>(Irr.CP)</sub>) for 4 weeks

#### *Blood sampling:*

Directly, after animals were anaesthised using diethyl ether, heparinized blood samples were collected from all animal groups after 2 and 4 weeks from the heart for determination of the following

#### *Biochemical parameters:*

SOD [9], GPx [10, 11] and CAT [12] (in packed RBCs) activities and plasma MDA [13], GSH (in packed RBCs) [14] and plasma NO (x) [15] concentrations. Plasma AST, ALT [16] and GGT [17] activities (as liver function tests), urea [18] and creatinine [19] concentrations (as kidney function tests).

#### *Statistical analysis:*

Statistical analysis was done using SPSS software version 15. The inter-group variation was measured by one way analysis of variance (ANOVA) followed by Post Hoc LSD test. Results were expressed as mean  $\pm$  SEM. The mean difference is significant at the 0.05 level [20].

### 3. RESULTS AND DISCUSSION

#### *Antioxidant parameters:*

The presented data in tables (1) revealed that, a highly significant decrease in SOD activity ( $p < 0.01$ ) after 2 weeks, a very highly significant decrease ( $p < 0.001$ ) in CAT and GPx activity and GSH content and a very highly significant increase in  $\text{NO}_{(x)}$  and MDA concentration after 2 and 4 weeks in TBM group when compared to control group (NTBM). These findings were in agreement with [21] who found that, the presence of tumor caused disequilibria of the antioxidant defense system. Moreover, [22] demonstrated that, lipid peroxidation level was significantly increased in blood, liver and tumor tissues of EAC mice when compared with control group. Also, our findings are in accordance with [23] who demonstrated that a decrease in blood GSH in circulation has been reported in several diseases including malignancies. Decline in SOD activity recorded in mice bearing Ehrlich carcinoma was also reported earlier by [24]. They postulated that the loss of Mn-SOD activity could be due to the loss of mitochondria which leads to a decrease in total SOD activity in different tissues of the tumor host. It seems that oxidative damage caused by decreased capacity for  $\text{H}_2\text{O}_2$  elimination is related to suppressed activity of CAT, as well as to suppressed direct antioxidant action of GSH. This is in agreement with the previous findings that CAT has a more significant role than GPx in protecting erythrocytes against oxidative stress [25-26].

Some investigators have reported a higher  $\text{NO}^{\cdot}$  synthase activity in tumors [40],

while some have reported a lower activity [27]. The obtained result supports the general observation that some malignancies are associated with an increased level of nitric oxide. In contrary, [28] who suggested that, there is a decrease rate of lipid peroxidation in liver tumor cell than normal liver cells.

Treatment with CP showed a non-significant increase in SOD activity and significant increase after 4 weeks in GPx activity and after 2 and 4 weeks in GSH content and CAT activity compared to TBM group. Furthermore, a significant decrease in MDA concentration was observed after 4 weeks. Also, significant decrease in  $\text{NO}_{(x)}$  concentration was reported after 2 and 4 weeks when compared with TBM group. But, treatment with Irr.CP revealed significant increase in SOD activity after 4 weeks compared to TBM and CP treated groups, in GPx activity after 4 weeks, in GSH content after 2 and 4 weeks compared to TBM group. Also, showed significant increase in CAT activity after 2 and 4 weeks compared to TBM group and after 4 weeks compared to CP treated group. Moreover, it indicated significant decrease in MDA concentration after 4 weeks compared to TBM group and CP treated group. Also, a significant decrease in  $\text{NO}_{(x)}$  concentration was observed after 2 and 4 weeks compared to TBM and after 2 weeks compared to CP. Our results are in harmony with [29], who reported that pectin could reduce MDA levels and increase SOD in aorta tissue in high fat diet fed rats. Also, [30] reported that, the addition of pectin to the cystine diet counteracted the activities of the total and Cu, Zn-superoxide dismutase, and of catalase in liver. Moreover, [31], reported that pectin extracted from citrus and grapefruit peel in laying hens diet increases blood serum SOD activity. Our results are disagreeing with [32] who reported that, erythrocyte SOD activity was not affected by pectin treatment in hypercholesterolemic rabbits. Moreover, our results are in agreement with [33], the

bioactivity of SOD and GSH-Px increased in all MCP-fed groups and the level of MDA decreased markedly in hyperlipidemic rats. It has been suggested that pectin interacts directly with oxidants and free radicals [34]. The antioxidant activity in pectin could be related to the high galacturonic acid content. It has been reported that a relatively low molecular weight and a high uronic acid content in polysaccharides appeared to increase the antioxidant activity [35] and this express the high antioxidant activity of modified citrus pectin (pectin degraded by irradiation) more than citrus pectin without degradation.

*Liver and kidney functions:*

The obtained results in tables (2) showed highly significant increase after 4 weeks in ALT activity and after 2 and 4 weeks in AST activity, furthermore, it revealed very highly significant increase in GGT activity in TBM group compared to control group. Meanwhile, a highly significant increase in urea and creatinine concentrations in TBM group compared to NTBM group. The results of the present work agreed with previous studies which have demonstrated that the level of the liver enzymes increased in serum of EAC-bearing mice indicating general toxicity that occurred due to cancer development [36]. The same results were observed by [37] in their study on the liver function in the assessment of head and neck cancer patients. The observed increase in serum urea level in tumor bearing mice are in agreement with the results reported by [38] who observed that, there was a significant increase in plasma urea concentration in tumor-bearing mice, he attributed such increase to the increase in urea production as a result of catabolic effect of tumor. As confirmed by [39] who suggested that, creatinine was decreased in tumor-bearing rats as the glomerular lesions progressed, associated with a rise in serum creatinine

level. Also, liver and kidney toxicity induced during tumor growth may be due to the excessive production of ROS that leads to oxidative damage [40]. It was previously observed that oxidative damage which appeared as increased lipid peroxidation and inhibition of GSH content, catalase and SOD activity, led to liver and kidney dysfunctions [41]. Our results revealed that treatment with CP revealed significant decrease in ALT and AST activities after 2 weeks and in urea and creatinine concentration after 4 weeks compared to TBM group. Meanwhile, treatment with Irr.CP showed significant decrease in ALT activity after 2 weeks compared to TBM group and in GGT activity after 4 weeks compared to TBM group and CP treated group. Also, it showed significant increase in urea and creatinine concentration after 4 weeks compared to TBM group. Similarly reported it was reported that, ALT, AST, GGT activities, serum urea and creatinine were significantly increased in positive control rat groups' administrated lead acetate [24]. Low and high esterified pectin significantly decreased the effect of LA on the tested parameters. Who added that, histopathological examination clearly indicated that high or low esterified pectin eliminated the harmful effect of LA on liver, kidney and brain tissues and showed no significant difference in serum ALT, AST, GGT activities, serum urea and creatinine in treated groups with high and low esterified pectin compared to normal control group.

#### **4. CONCLUSION AND RECOMMENDATIONS**

We recommended using citrus pectin and irradiated citrus pectin in our food as prophylactic and preventive for many diseases and as adjuvant therapy in cancer thereby.

Table 1 Effect of treatment with citrus pectin and irradiated citrus pectin on SOD, GPx, CAT activities, GSH, MDA and NO (x) (µM/L) concentrations in Ehrlich carcinoma bearing female mice and their control during 4 weeks:

Parameters	Group	weeks	NTBM	TBM	TBM <sub>(CP)</sub>	TBM <sub>(Irr.CP)</sub>
SOD(U/ml)		2	3.34±0.09	2.58 <sup>a</sup> ±0.015	2.91 <sup>d</sup> ±0.012	3.04 <sup>d</sup> ±0.017
		4	3.22±0.08	2.60±0.06	2.70 <sup>f</sup> ±0.041	3.09 <sup>ab</sup> ±0.06
GPx(mg/dl)		2	0.78±0.006	0.64 <sup>11</sup> ±0.0007	0.62 <sup>d</sup> ±0.0038	0.64±0.0038
		4	0.78±0.0007	0.61 <sup>11</sup> ±0.0010	0.67 <sup>e</sup> ±0.011	0.65 <sup>g</sup> ±0.016
GSH(mg/dl)		2	137.74±1.64	109.96 <sup>11</sup> ±2.13	145.42 <sup>g</sup> ±3.25	153.49 <sup>h</sup> ±5.58
		4	140.67±4.22	109.96 <sup>11</sup> ±6.14	154.43 <sup>g</sup> ±4.65	154.11 <sup>h</sup> ±5.68
CAT (µM/ml)		2	0.595±0.014	0.350 <sup>11</sup> ±0.017	0.462 <sup>g</sup> ±0.037	0.433 <sup>g</sup> ±0.022
		4	0.610±0.007	0.541 <sup>11</sup> ±0.004	0.479 <sup>g</sup> ±0.004	0.574 <sup>h</sup> ±0.018
MDA(µM/ml)		2	91.740±1.680	137.73 <sup>11</sup> ±4.878	120.405 <sup>g</sup> ±4.029	128.905 <sup>g</sup> ±7.380
		4	92.685±2.479	129.888 <sup>11</sup> ±5.314	108.408 <sup>g</sup> ±3.947	88.072 <sup>h</sup> ±3.881
NO <sub>(x)</sub> (µM/L)		2	20.960±0.768	31.515 <sup>11</sup> ±0.816	31.675 <sup>11</sup> ±0.692	28.045 <sup>h</sup> ±0.799
		4	22.073±0.545	33.000 <sup>11</sup> ±0.608	18.980 <sup>g</sup> ±1.022	18.243 <sup>g</sup> ±1.802

Non-significant (N.S): p>0.05; Significant: \*p<0.05; highly significant: \*\* p<0.01; very highly significant: \*\*\*p<0.001 from NTBM. a, significant from TBM group p<0.05. b, significant from TBM<sub>(CP)</sub> group p<0.05. c, significant from TBM<sub>(Irr.CP)</sub> group p<0.05

Table 2 Effect of treatment with citrus pectin and irradiated citrus pectin on blood ALT , AST , GGT activities, Urea and Creatinine concentrations in Ehrlich carcinoma bearing female mice and their control during 4 weeks:

Parameters	ALT(U/L)		AST(U/L)		GGT(U/L)		Urea(mg/dl)		Creatinine(mg/d)	
	2wks	4wks	2wks	4wks	2wks	4wks	2wks	4wks	2wks	4wks
NTBM	50.51 ±2.80	51.64 ±0.71	96.50± 4.99	102.32 ±2.64	31.01 ±2.22	31.81 ±0.78	29.65± 2.52	29.10 ±0.67	0.79 ±0.07	0.78±0.017
TBM	56.45 ±4.76	66.43 <sup>**</sup> ±5.14	130.45 <sup>***</sup> ±6.16	123.84 <sup>**</sup> ±4.64	52.62 <sup>***</sup> ±0.73	57.56 <sup>***</sup> ±0.39	28.85 ±2.84	33.20 <sup>**</sup> ±1.24	0.77 ±0.06	0.89 <sup>**</sup> ±0.32
TBM <sub>(CP)</sub>	71.07 <sup>a</sup> ±1.97	60.74 ±3.19	100.39 <sup>a</sup> ±1.42	119.53 ±1.37	52.72 ±1.78	55.05 <sup>c</sup> ±1.05	26.15 ±0.35	26.15 <sup>a</sup> ±0.35	0.70 ±0.09	0.78 <sup>a</sup> ±0.13
TBM <sub>(Irr.CP)</sub>	69.14 <sup>a</sup> ±0.88	69.89 ±3.07	118.30 ±11.30	120.10 ±6.91	50.80 ±1.77	50.09 <sup>bc</sup> ±2.14	28.90 ±1.515	28.10 <sup>a</sup> ±0.84	0.77 ±0.04	0.75 <sup>a</sup> ±0.23

Non-significant (N.S): p>0.05; Significant: \*p<0.05; highly significant: \*\* p<0.01; very highly significant: \*\*\*p<0.001 from NTBM. a, significant from TBM group p<0.05. b, significant from TBM<sub>(CP)</sub> group p<0.05. c, significant from TBM<sub>(Irr.CP)</sub> group p<0.05.

## 5. REFERENCES

- Lingwood, R.; Boyle, P., Milburn, A. T., Ngoma, Arbuthnott, J., McCaffrey, R., Kerr, S., and Kerr, D. 2008. The challenge of cancer control in Africa. *Nat. Rev. Cancer.* **8**: 398–403.
- Om-Ali, Y.K., Tarek, A.S., and Mohamed, F.S. 2003. Protective role of Egyptian propolis against tumor in mice. *Clinica Chimica Acta. J.*, **338**:11-16.
- Nangia-Makker, P.; Hogan, V., Honjo, Y., Baccarini, S., Tait, L., Bresalier, R. and Raz, A. 2002. Inhibition of human cancer cell growth and metastasis in nude mice by oral intake of modified citrus pectin. *J. Natl. Cancer Inst.* **94**:1854–1862.
- Azemar, M.; Hildenbrand, B., Haering, B., Heim, M.E. and Unger, C. 2007. Clinical Benefit in Patients with Advanced Solid Tumors Treated with Modified Citrus Pectin: A Prospective Pilot Study. *J. Clinical Medicine: Oncology.* **1**:73-80.
- Kang, H.J.; Jo, C., Kwon, J.H., Son, J.H., An, B.J. and Byun, M.W. 2006. Antioxidant and cancer cell proliferation inhibition effect of citrus pectin

## Anti-tumor activity of irradiated citrus pectin

- oligosaccharide prepared by irradiation. *J. Med. Food.* **9**: 313-320.
6. Cho, M.; Kim, B.Y. and Rim J. H. 2003. Degradation of alginate solution and powder by  $\gamma$ -radiation. *J. Food Eng. Prog.*, **7**:141-145.
  7. Chen, C.H.; Sheu, M.T., Chen, T.F., Wang, Y.C., Hou, W.C., Liu, D.Z., Chung, T.C. and Liang, Y.C. 2006. Suppression of endotoxin induced proinflammatory responses by citrus pectin through blocking LPS signaling pathways. *J. Biochem. Pharmacol.* **72**: 1001-1009.
  8. Attari, F.; Sepehri, H., Delphi, L. and Goliae, B. 2009. Apoptotic and necrotic effects of pectic acid on rat pituitary GH3/B6 tumor cells. *Iranian Biochemical J.*, **13**(4):163-170.
  9. Minami, M. and Yoshikawa, H. 1979. A simplified assay method of superoxide dismutase activity for clinical use. *Clin. Chim. Acta* **92**:337-342.
  10. Gross, R. T.; Bracci, R., Rudolph, N., Schroeder, E., and Kochen, J. A. 1967. Hydrogen peroxide toxicity and detoxification in the erythrocytes of newborn infants. *J. Blood.* **29**: 481-493.
  11. Necheles, T. F., Boles, T., and Allen, D. M. 1968. Erythrocyte glutathione peroxidase deficiency and hemolytic diseases of the newborn infant, *J. Ped.* **72**:319-324.
  12. Sinha, A.K. 1972. Colorimetric assay of catalase. *J. Anal Biochem.* **47**: 389-394.
  13. Yoshioka, T.; Kawada, K.; Shimada, T and Mori, M. 1979. Lipid peroxidation in maternal and cord blood and protective mechanism against activated oxygen toxicity in the blood. *Am. J. Obsterics. Gynecology*, **135**: 372-376.
  14. Beutler, E.; Duron, O. and Kelly, B. M. 1963. Improved method for the determination of blood glutathione. *J. Lab. Clin. Med.*, **61**: 882-888.
  15. Chen, C.H.; Sheu, M.T., Chen, T.F., Wang, Y.C., Hou, W.C., Liu, D.Z., Chung, T.C. and Liang, Y.C. 2006. Suppression of endotoxin induced proinflammatory responses by citrus pectin through blocking LPS signaling pathways. *J. Biochem Pharmacol.* **72**: 1001-1009.
  16. Miranda, K.M.; Espey, M.G. and Winl, D.A. 2001. A rapid, simple spectrophotometric method for simultaneous detection of nitrate and nitrite. *Nitric Oxide Boil. Chem.* **5**: 62-71.
  17. Reitman, S. and Frankel, S. 1957. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *Am J Clin Pathol.* **28**: 56-63.
  18. Szasz, G. 1969. A kinetic photometric method for serum Gamma glutamyle transpeptidase. *J. Clin chem.*, **15**: 124-136.
  19. Palton, C.J. and Crouch, S.R. 1977. Spectrophotometric and kinetics investigation of the Berthelot reaction for the determination of ammonia. *J. Anal Chem.* **49**: 464-469.
  20. Henry, R.J.; Cannon, D.C. and Winkelman, J.W. 1974. Clinical chemistry: Principles and technics. *Hagerstown, Maryland: Harper and Row. Pp.* 1106.
  21. Snedecor and Cochran 1969. Statistical methode 6th ed. The Iowa State Univ., Press, Iowa, USA.
  22. Kumaraguruparan R, Subapriya R, Kabalimoorthy J and Nagini S. 2002. Antioxidant profile in the circulation of patients with fibroadenoma and adenocarcinoma of the breast. *Clin Biochem.* **35**: 275-279.
  23. Hayat, M.S. 2001. Effect of inositol hexaphosphate (IP6) on the activity of antioxidant defense system in mice loaded with solid tumor. *Egyptian Journal of biochemistry and molecular biology.* **24**:137-153.
  24. Saygili, E. I.; Akcay, T., KonuKoglu, D. and Papilla, C. 2003. Cgdem. Glutathine and glutathione-related enzymes in colorectal cancer patients. *J. Toxicol. Environ. Health.*, **66**: 411-415.
  25. Sahu, S.K.; Oberley, L.W., Stevens, R.H. and Riley, E.F. 1977. Superoxide dismutase activity of Ehrlich ascities tumor cells. *J Natl Cancer Inst.* **58**: 1125-1128.
  26. Gaetani, G. F.; Ferraris, A.M., Rolfo, M., Mangerini, R., Arena, S. and Kirkman, H.N. 1996. Predominant role of catalase in the disposal of hydrogen peroxide within human erythrocytes. *Blood.* **87**: 1595-1599.

27. Muller, S.; Riedel, H.D. and Stremmel, W. 1997. Direct evidence for catalase as the predominant H<sub>2</sub>O<sub>2</sub>-removing enzyme in human erythrocytes. *Blood*. **90**: 4973-4978.
28. Janson, O.T.; Morcos, E., Brundin, L., Bergerheim, U. S. R., Adolfsson, J. and Wiklund, N.P. 1998. Nitric oxide synthase in human renal cell carcinoma. *J. Urol.*, **160**: 556-560.
29. Cheeseman, K.H.; Emery, S.P.; Maddix, S.P.; Slater, T.F.; Burton, G.W. and Ingold, K.U. 1988. Studies on lipid peroxidation in normal and tumor tissue. *Biochem. J.* **250**: 247-252.
30. Xiaojian, S.; Xiuzhen, Z. and Huiyun, W. 1997. The effect of pectin on lipid and lipid peroxide levels in aorta, heart and brain tissues of rats. *ACTA Acadimae Medicinae Qingdao*. 1997-2001.
31. He, G. and Aoyama, Y. 2003. Effects of adding some dietary fibers to a cystine diet on the activities of liver antioxidant enzymes and serum enzymes in rats. *Biosci Biotechnol Biochem.* **67**: 617-621.
32. Lien, T.; Yeh, H.S. and Su, W.T. 2008. Effect of adding extracted hesperetin, naringenin and pectin on egg cholesterol, serum traits and antioxidant activity in laying hens. *Archives of Animal Nutrition.* **62**: 33-43.
33. Ismail, M.F.; GAD, M.Z., and Hamdy, M.A. 1999. Study of the hypolipidemic properties of pectin, garlic and ginseng in hypercholesterolemic rabbits. *Pharmacolog. Research J.* **39**: 157-166.
34. Ai-zhen, Z. and Li-fang, Z. 2007. Effects of Modified Citrus Pectin on Lipid Peroxidation in Hyperlipidemic Rats. *Zhejiang Journal of Preventive Medicine.* 2007-2012.
35. Khasina, E.I.; Kolenchenko, E.A., Sgrebneva, M.N., Kovalev, V.V. and Khotimchenko, Y.S. 2003. Antioxidant activities of low etherified pectin from the seagrass *Zostera marina*. *Russ. J. Mar. Biol.* **29**:259-261.
36. Chen, H.X.; Zhang, M. and Xie, B.J. 2004. Quantification of uronic acids in tea polysaccharides conjugates and its antioxidant properties. *J. Agric. Food Chem.* **52**:3333-3336.
37. Pal, S.; Bhattacharyya, S., Choudhuri, T., Datta, G.K., Das, T. and Sa, G. 2005. Amelioration of immune cell number depletion and potentiation of depressed detoxification system of tumor-bearing mice by curcumin. *Cancer Detect Prev.* **29**: 470-478.
38. Korver, K.D.; Graham, S.M., Hoffman, H.T., McCulloch, T. and Funk, G.F. 1995. Liver function studies in the assessment of head and neck cancer patients. *Head Neck.* **17**: 531-534.
39. Hussein, S.A. and Azab, M.E. 1997. Effect of insulin treatment on some metabolic changes on experimentally induced tumor in female mice. *The Egyptian J. of Biochemistry.* **15**: 61-80.
40. Kawaguchi, H.; Itoh, K., Mori, H., Hayashi Y and Makino, S. 1991. Renal pathology in rats bearing tumor-secreting growth hormones. *Pediatr Nephrol- Jul* .**5**: 533-8.
41. Borges, L.P.; Nogueira, C.W., Panatieri, R.B., Rocha, J.B. and Zeni, G. 2006. Acute liver damage induced by 2-nitropropane in rats: effect of diphenyl diselenide on antioxidant defenses. *Chem Biol Interact.* **160(2)**:99-107.
42. El-Nahal, D. M. 2010. Effect of using pectin on lead toxicity. *J. American Science.* **6**: 441-554.



## التقييم الكيميائي الحيوي للنشاط المضاد للأورام لبكتين الموالح المشع

أميمه أحمد رجب أبو زيد<sup>1</sup> - احمد إبراهيم البطل<sup>2</sup> - عفت سليمان إسماعيل<sup>1</sup>

<sup>1</sup> قسم الكيمياء الحيوية والإكلينيكية- كلية الطب البيطري بمشهر - جامعة بنها، <sup>2</sup> قسم البحوث الدوائية الإشعاعية، شعبة التكنولوجيا الحيوية، المركز القومي لبحوث وتكنولوجيا الإشعاع، هيئة الطاقة الذرية.

### الملخص العربي

يعد مرض السرطان من اهم المشكلات الصحية على مستوى العالم حيث انه يعتبر من اكثر المسببات للوفاة فى الدول النامية والدول المتقدمة، وقد أثبتت الدراسات أن من 80% إلى 90% من حالات السرطان سببها التلوث البيئي كما أنه يصعب التحكم فيها وعلى الرغم من نجاح الجراحة في استئصال بعض الأورام إلا أن انتشار المرض أو ظهوره مرة أخرى مشكلة توجد في معظم الأورام. ومن ثم تكاثفت الجهود العالمية لاستخلاص مركبات طبيعية مضادة للسرطان لها تأثير قوى وفعال ضد الأورام، ومن هنا توجهت الأنظار إلى استخدام مستخلصات طبيعية ليس لها أي تأثير جانبي على المرضى عند العلاج. وهو هدف هذا البحث وهو التقييم الكيميائي الحيوي للنشاط المضاد للأورام لبكتين الموالح المشع. أجريت هذه الدراسة على عدد 60 من إناث الفئران السويسرية البيضاء والتي تزن من 20-25 جم منها 45 فارا تم حقنهم ب 10 X 2.5<sup>6</sup> من خلايا ايرلخ السرطانية في فخذ الفأر وتركت جميعا حتى وصل قطر الورم إلى 10م3 ثم تم تقسيم الفئران إلى 4 مجموعات متساوية (15 فأر لكل مجموعة) كما يلي: المجموعة الأولى: مجموعة ضابطة سليمة (سالبة) تم تجريعها بالمحلول الملحي. المجموعة الثانية: مجموعة ضابطة موجبة وهى تحمل الورم تم تجريعها بالمحلول الملحي لمدة 4 أسابيع . المجموعة الثالثة: مجموعة حاملة للورم وتم تجريعها ببكتين الموالح (3.3 جم/كجم من وزن الحيوان يوميا) لمدة 4 أسابيع. المجموعة الرابعة: مجموعة حاملة للورم وتم تجريعها ببكتين الموالح المشع(3.3 جم/كجم من وزن الحيوان يوميا) لمدة 4 أسابيع. تم تجميع عينات الدم لقياس بعض الدلالات الحيوية بها مثل قياس بعض مضادات الأكسدة مثل المألون ثنائي الالدهيد، أكسيد النيتريك ، مستوى الجلوتاثيون ، ونشاط كل من إنزيم الجلوتاثيون بيروكسيداز ، السوبر اوكسد ديسميونيز ، وأيضا تم قياس كلا من إنزيمات الكبد الألائين والاسبرتيت الناقلين للأمين وجاما جلوتاميل ترانسفيريز وأيضا قياس مستوى البولينيا والكرياتينين. وأضحنت النتائج زياده معنويه في وظائف الكبد والكلى بالإضافة إلى زيادة معنوية فى تركيز المألون ثنائي الالدهيد و اكسيد النيتريك ونقص معنوي في نشاط إنزيم الكاتلاز والسوبر اوكسيد ديسميونيز والجلوتاثيون بيروكسيداز وتركيز الجلوتاثيون في الفئران الحاملة للورم السرطاني بالمقارنة بالمجموعة الضابطة وحدث تغير في هذه النتائج إلى الأفضل بعد التجريع ببكتين الموالح وبكتين الموالح المشع. لذا نوصى بتناول بكتين الموالح وبكتين الموالح المشع كعامل وقائي أو كعلاج مساعد في علاج الأورام السرطانية.

(مجلة بنها للعلوم الطبية البيطرية: عدد 23 (1)، يونيو 2012: 11-18)